

Association of TP53 Polymorphism in Lung Cancer Patients and Healthy People in Bangladesh

¹Shahin Mahmud, ¹Saurav Dey Sarker, ²Shoaib Mahmud Shaon, ⁴Abu Saim al Salauddin, ³Md. Khaja Foyasal, ³Rafsan Zani Tanvir, ¹Masum Parvez, ¹Hasibul Haque Rakib, ¹Binita Shome, ¹Md. Shariful Islam *

¹Department of Biotechnology and Genetic Engineering, Faculty of Life science, MawlanaBhashani Science and Technology University, Tangail-1902, Bangladesh

²Department of Biochemistry and Molecular Biology, Jahangirnagar University, Savar-1342, Bangladesh

³Department of Biotechnology and Genetic Engineering, Faculty of Applied Science and Technology, Islamic University, Kushtia-7003, Bangladesh

⁴Department of Microbiology and Hygiene, Faculty of Veterinary Science, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh

Corresponding author: Md. Shariful Islam, Department of Biotechnology and Genetic Engineering, Faculty of Life science, MawlanaBhashani Science and Technology University, Tangail-1902, Bangladesh.

Corresponding author mail: sharifbge@gmail.com

Abstract: Lung cancer is one of the most common and lethal cancer. It's most important etiologic factor is tobacco smoking. The p53 tumor suppressor protein is important in cell-cycle control, apoptosis, and DNA repair. Polymorphisms in TP53 have been associates with inherited cancer susceptibility. The association of p5 codon 72 polymorphisms with lung cancer has been investigated by several scientific groups with controversial results. However, the genotype distribution of p53 codon 72 polymorphisms as well as the association of this polymorphism with lung cancer risk remains undefined in the Bangladesh population. It has been reported that the genotype distribution of p53 codon 72 polymorphism of lung cancer patients and non-cancer controls and association of demographic characteristics with lung cancer. In a case control study, we frequently matched (by age, sex, and smoking) lung cancer patients and control subjects. TP53 Polymorphisms were determined by restriction fragment length Polymorphisms (RFLP) analysis of PCR amplified exon 4 of p53 gene. Nearly one-third of total lung cancer incidence in Bangladesh was squamous cell carcinoma subtype and increased frequency of Arg/Arg and Pro/Pro genotypes were associated with non-small cell lung cancer (NSCLC). The absence of Pro/Pro genotypes in non-smoker and association with low pack years smoking status suggested that Pro/Pro genotypes was more frequently affected by tobacco carcinogens, p53 polymorphism may be associated with increased lung cancer risk and may affect the function of p53 gene. The aim of this present review is to Investigation of the effect of smoking and TP53 polymorphism in developing lung cancer and Study the association of TP53 polymorphism in lung cancer patient and healthy people in Bangladesh.

Keywords: p53 Tumor suppressor protein, Tobacco smoking, Restriction fragment length Polymorphisms, Lethal cancer, Lung cancer, Carcinoma

1. Introduction

Lung cancer involves malignant proliferation of the epithelial lining of the lower respiratory tract. It is one of the most common forms of malignancy leading to the major cause of cancer related deaths across the world (Parkinet *al.*,2001) and also in Bangladesh (CRR, 2009) Lung cancer has been the most common cancer in the world since 1985 and the leading cause of cancer death. Worldwide it is by far the most common cancer of men and increasingly being recognized in Bangladesh. An estimated 1.61 million people across the world were diagnosed with lung cancer in 2008,

accounting for 13% of the total. More than half (55%) of the cases occurred in the developing world (Ferlay *et al.*, 2008). Lung cancer incidence is more than double in men than in women worldwide (rate ratio 2.5: 1.0) (Ferlay *et al.*, 2008). Lung cancer was one of the common cancers among the patients who attended National Institute of Cancer Research and Hospital (NICRH), Dhaka, Bangladesh. A total of 3,209 lung cancer patients attended during 2005- 2007, of them 86% (2,763) were males. The number of lung cancer patients was increasing year by year; there were 902 lung cancer patients in 2005, 1,076 in 2006 and 1,231 in 2007(CRR. 2009). According to the latest who data

published in April 2011 lung cancers death reached 18,124 or 1.89% of total deaths. The age adjusted death rate is over 20.29 per 100,000 of population rank a #59 in the world.

Lung cancer is mainly two types. One type is small cell lung carcinoma, where the cancer cells contain dense neurosecretory granules (vesicles containing neuroendocrine hormones), which give this tumor an endocrine/paraneoplastic syndrome association. Most cases arise in the larger airways (primary and secondary bronchi) (Behzadi et al., 2009). This cancer grows quickly and spread early in the course of the disease. At presentation around 60–70% of this type of cancer has metastatic disease. This type of lung cancer is strongly associated with smoking (Hogan, 1999). The other type is non-small cell lung carcinoma (NSCLC), which is more common, accounting for approximately 80 percent of all lung cancer cases (Dayem, 2002). Further, non-small cell lung carcinoma (NSCLC) is divided into three subtypes, Adenocarcinoma, Squamous cell carcinoma and large cell carcinoma. Adenocarcinoma is more common in women and non-smokers; this type of NSCLC arises from the terminal bronchioles or alveolar walls (Dayem, 2002). Most cases of adenocarcinoma are associated with passive smoking or people who have smoked fewer than 100 cigarettes in their lifetimes ("never-smokers") (Hogan, 1999). It grows more slowly than the Squamous cell carcinoma. It spreads to other parts of the body at an early stage, and accounts for 40 percent of all cases (Dayem, 2002). Squamous cell carcinoma is associated with smoking and found mostly in men and older people of both sexes. It accounts for 30 to 35 percent of all NSCLC (Dayem, 2002). They typically occur close to large airways. A hollow cavity and associated necrosis are commonly found at the center of the tumor (Rahaman et al., 2011). Large cell carcinoma accounts for 5 to 15 percent of all case (Dayem, 2002). These are so named because the cancer cells are large, with a lot of cytoplasm, large nuclei and conspicuous nucleoli (Rahaman et al., 2011).

1.2 TP53 Gene

The human *TP53* tumor suppressor gene, located on chromosome 17p13, is involved in several central cellular processes, including gene transcription, DNA repair, cell cycling, apoptosis and genomic stability. The *TP53* is one of the most frequently mutated genes in human cancers. It is reported that approximately half of all cancers have inactivated *TP53* (Robles and Harris, 2010). To date, several polymorphisms in the wild type *TP53* gene locus have been described. Among them the codon72 polymorphism on the exon 4 of the *TP53* gene, that have either arginine (CGC) or proline (CCC), has been reported to be associated with bladder and lung cancer (Carbone, 1991). The association of p53 codon72 polymorphism has been studied in lung carcinoma by several authors.

However, the results are contradictory. While some authors reported the frequency of Pro/Pro genotype to be higher in lung cancer (Fan et al., 2000; Kawajiri et al., 1993; Wang et al., 1999; Weston et al., 1992b). Papadakis et al., (2002) observed an increased frequency of Arg/Arg genotype in advanced lung cancer cases (Pandima et al., 2010) indicated that p53 Arg genotypes are more susceptible to lung cancer in India. (Pandima et al., 2010) also showed that the frequency of Arg/Arg is higher in smokers. The association p53 codon72 polymorphism with lung cancer has not ever been studied in Bangladesh. Therefore, we are intending to study the association. Cancer caused about 13% of all human death worldwide (7.9 million). Rates are rising as more people live to an old age and as mass lifestyle changes occur in the developing world. (Jemal, A et al., 2011)

1.3 Lung Cancer in Bangladesh

Lung cancer is the most commonly diagnosed cancer (17% of the total new cancer cases) and leading cause of death in males (23 % of the total cancer deaths) whereas it is the fourth most commonly diagnosed cancer (8.5 %) and the second leading cause of cancer death (12.8 %) in female (Jemalet al., 2011 and Ferlay et al., 2008). In Bangladesh it is also the most prevalent cancer in male by considering incidence and mortality (24.9 % of incidence and 28.6 % of mortality) where as it is the fourth most prevalent cancer by considering incidence and second leading cause of cancer related death in females (5.6 %) (Ferlay et al., 2008) According to WHO (2007) total lung cancer cases in Bangladesh were 196,000 among those age 30 years and above (Haque, 2011 and WHO, 2007). New lung cancer cases in Bangladesh were 20,904 and total mortality were 19,393 in 2010 (Ferlay et al., 2008) The annual number of new cases and mortality are estimated to rise to 43,048 and 40,252, respectively, by 2030 (Ferlay et al., 2008) In Bangladesh, Lung cancer incidence was the highest among commonly occurred cancers (16.7% of the total cancer cases) (Cancer Registry Report of the NICRH, 2005-2007) (CRR, 2009). In the similar fashion, lung cancer was the most common incident form of cancer in 2006 (16.4% of the total cases) and 2007 (17.3% of the total cases) in Bangladesh (CRR, 2009).

1.4 Causes of Lung Cancer

Lung cancer is most often a “multifactorial” disease. There are many lung cancer causes, in addition to the well-known link with smoking. An overview of common lung cancer causes includes:

1.5 Population Characteristics

Lung cancer risk increases with age in both smokers and never smokers. The cancer is sufficiently rare in people under the age of 40, especially among never smokers (Samet et al., 2009). Women who have never smoked are more

likely to develop lung cancer than men who have never smoked (Patel *et al.*, 2004).

1.6 Lifestyle

Smoking, particularly of cigarettes, is by far the main contributor to lung cancer. The incidence of lung cancer is strongly correlated with cigarette smoking, with about 90% of lung cancers arising as a result of tobacco use (Biesalski *et al.*, 1998). Tobacco smoke contains over 60 known carcinogenic compounds, Tobacco smoke contains over 4,000 chemical compounds, many of which have been shown to be cancer-causing or carcinogenic. The carcinogens in tobacco smoke are chemicals known as radioisotopes from the radon decay sequence, nitrosamine, and benzopyrene (Sopori, 2002). Additionally, nicotine appears to depress the immune response to malignant growths in exposed tissue (Sopori, 2002). Across the developed world, 91% of lung cancer deaths in men during the year 2000 were attributed to smoking (71% for women) (Ezzati and Lopez 2004). In the United States, smoking accounts for 80–90% of lung cancer cases (Hogan, 1999). The risk of lung cancer increases with the number of cigarettes smoked and the time over which smoking has occurred; doctors refer to this risk in terms of pack-years of smoking history (the number of packs of cigarettes smoked per day multiplied by the number of years smoked) (Hecht, 2003 and Sopori, 2002).

Passive smoking or the inhalation of tobacco smoke by nonsmokers, who share living or working quarters with smokers, also is an established risk factor for the development of lung cancer. Research has shown that nonsmokers who reside with a smoker have a 24% increase in risk for developing lung cancer when compared with nonsmokers who do not reside with a smoker (Taylor *et al.*, 2007). An estimated 3,000 lung cancer deaths that occur each year in the U.S. are attributable to passive smoking (Taylor *et al.*, 2007).

A higher intake of foods, such as salads, is associated with a lower risk of developing lung cancer. An excess intake of alcohol may raise the risk of lung cancer in smokers (Samet *et al.*, 2009).

1.7 Environmental Causes

Many environmental exposures can contribute to lung cancer risk in addition to smoking, and like smoking, many of these are avoidable if we are aware of them. You can reduce your risk by doing things as simple as testing your home for radon and using an appropriate mask when working with certain chemicals. Some of the most common environmental causes of lung cancer include:

1.8 Radon

Exposure to radon in the home is the second-leading cause of lung cancer and the leading cause in nonsmokers. Radon is a radioactive colorless

and odorless gas that is produced by the natural decay of uranium, found in the Earth's crust. The radiation decay products ionize genetic material, causing mutations that sometimes turn cancerous (Alberg, 2007). It is estimated 12% of lung-cancer deaths attributable to radon gas, or about 20,000 lung-cancer-related deaths annually in the U.S., making radon the second leading cause of lung cancer in the U.S. The U.S. Environmental Protection Agency estimates that one out of every 15 homes in the U.S. contains dangerous levels of radon gas (Schmid *et al.*, 2010).

1.9 Asbestos

Asbestos fibers are silicate fibers that can persist for a lifetime in lung tissue following exposure to asbestos. Asbestos can cause a variety of lung diseases, including lung cancer. There is a synergistic effect between tobacco smoking and asbestos in the formation of lung cancer (Hubaux *et al.*, 2012). Exposure to asbestos is responsible for the majority of mesothelioma, a rare tumor involving the lining of the lung (which is different from lung cancer) (Davies *et al.*, 2010).

1.10 Air Pollution

Outdoor air pollution, especially in urban areas, appears to raise the risk of lung cancer. Outdoor air pollution is estimated to account for 1–2% of lung cancers (Alberget *et al.*, 2010). Atomic bomb survivors have an elevated risk of developing lung cancer, as do those who have undergone radiation therapy for other types of cancer, such as Hodgkin's disease. Fine particulates (PM_{2.5}) and sulfate aerosols, which may be released in traffic exhaust fumes, are associated with slightly increased risk (Chen *et al.*, 2008).

1.11 Occupational Causes

A number of occupations or occupational exposures are established or suspected risk factors for lung cancer. The International Agency for Research on Cancer has identified 12 occupational exposure factors as being carcinogenic to the human lung (aluminum production, arsenic, asbestos, bis-chloromethyl ether, beryllium, cadmium, hexavalent chromium, coke and coal gasification fumes, crystalline silica, nickel, radon, and soot). Diesel exhaust has been classified as probably carcinogenic to humans (Steenland *et al.*, 1996). Welding of stainless steel is possibly associated with an increased risk of lung cancer, but the evidence for a carcinogenic effect of welding for other materials is weak (Sjogren *et al.*, 1994 and Moulinet *et al.*, 1997). However, the potential confounding effect from asbestos exposure may not have been fully controlled in many studies (Moulinet *et al.*, 1997). The carcinogenic effect of certain polycyclic aromatic hydrocarbons (PAHs), especially benzo(a)pyrene, is well documented (International Agency for Research on Cancer, 1983). The lung cancer excess in occupations involving high exposure to combustion products is

often attributed to exposure to PAHs, possibly in combination with exposure to particles (Gustavsson *et al.*, 1998). While dermal exposure to low grade mineral oils is carcinogenic to humans, there is less evidence for a cancer hazard from inhalation of oil mist (Tolbert *et al.*, 1997). Occupational exposure to carcinogens is estimated to be responsible for 13 to 29% of lung cancers in men (Samet *et al.*, 2009).

1.12 Genetics

In general, a genetic predisposition to lung cancer does not mean someone will develop lung cancer. It means they are more likely to develop lung cancer, especially when combined with other risk factors. Having a first-degree family member (parent, sibling or child) with lung cancer roughly doubles the risk of developing lung cancer. This risk is more for women and less for men and stronger in nonsmokers than smokers. Having a second-degree relative (an aunt, uncle, niece or nephew) with lung cancer raises the risk by around 30% (Kern *et al.*, 2008). In relatives of people with lung cancer, the risk is increased 2.4 times. This may be due to genetic polymorphisms (Kern *et al.*, 2008). Studies vary in the types of lung cancers that have the greatest hereditary component, but those with non-small cell lung cancers, especially lung adenocarcinoma are more likely to have a family history of lung cancer than those with small cell lung cancers (Kern *et al.*, 2008). Current and former smokers are at greatest risk, but lung cancer does occur among non-smokers, with varying rates across countries (Jema *et al.*, 2008). The association between cigarette smoking and increased risk of lung cancer is now undisputed. Despite this, less than 20% of smokers develop lung cancer, suggesting that the effect of tobacco smoke exposure is modified by other variables, including individual susceptibility (United States Department of Health and Human Services, 2004). The search for a gene or genes associated with susceptibility is still nascent. Genome wide association studies have independently reported chromosomal region 15q24-25.1, which contains nicotinic acetylcholine receptor sub-unit genes, to be associated with increased risk of lung cancer in ever smokers (Amos *et al.*, 2008; Hung *et al.*, 2008 and Thorgeirsson *et al.*, 2008). These findings have been replicated among individuals with a family history of lung cancer, and the relative risk of lung cancer associated with markers in this region are much higher for familial cases compared to the relative risk observed among sporadic cases (Liu *et al.*, 2008). Linkage analysis in families with aggregation of lung cancer also described a region on chromosome 6q23-25 associated with risk of lung cancer (Bailey *et al.*, 2004). The clinical significance of these findings is still unclear. In the meantime, lung cancer risk models using epidemiologic data have been developed, and the most parsimonious models for both ever and never

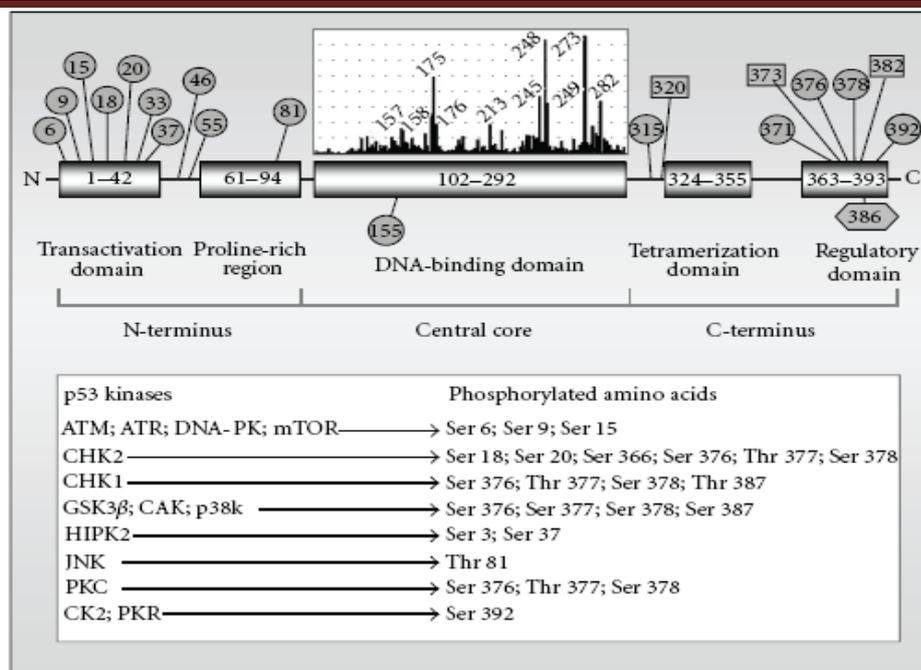
smokers include a family history of cancer variable (D'Amelio *et al.*, 2010).

1.13 p53 Protein

The p53 protein was initially identified as an oncogene because of its high expression in tumours. Soon after its discovery, it was found that the sequence of tumour-derived p53 differs from that originating from normal tissue. This led to the conclusion that the normal p53 protein acts as tumour suppressor rather than an oncogene. Meanwhile, p53 has become one of the most studied tumour suppressors, because of its central role in counteracting cellular transformation. Interestingly, two p53-related genes, p63 and p73, were identified in 1997 (Aramayo *et al.*, 2011; B. P. *et al.*, 2006). Both proteins share homology with p53 but differ in their biological functions from p53. In certain conditions, however, also p63 and p73 can function as tumour suppressors. The p53 protein is a transcription factor that specifically binds sequences as a tetramer to target (Bode and Dong 2004). The wt-p53 protein consists of 393 amino acids (aa) that constitute five functional domains, including two N-terminal transactivation domains (TAD1: aa 1–42, TAD2: aa 43–63) that are followed by a proline-rich domain (aa 64–92), the central DNA-binding domain (DBD: aa 102–292), the C-terminal tetramerization domain (aa 326–356) and basic domain (aa 364–393) (Aramayo *et al.*, 2011 and B. P. *et al.*, 2006) (Figure 1)

Figure 1: Structure and Post-transcriptional Modification of the Human p53 Protein. The insert vertical bars above the DNA-binding domain illustrate the distribution and the prevalence of point mutations found in p53 in human cancers. The most frequently mutated codons (“hotspot” codons) are identified. Several proteins that phosphorylate p53 and the target amino acids of p53 are also shown in the box. ATM, ataxiatelangiectasia mutated; ATR, ataxiatelangiectasia and RAD3 related; DNA-PK, DNA-dependent protein kinase; mTOR; mammalian target of rapamycin; CHK1 and 2, checkpoint kinases 1 and 2; p38k, p38 kinase; GSK3β, glycogen synthase kinase-3 beta; HIPK2, homeodomain interacting protein kinase 2; JNK, Jun-NH(2)-terminal kinase; PKC, protein kinase C; CK2, casein kinase 2; PKR, protein kinase R (Mirzayans *et al.*, 2012).

The N-terminal acidic transcriptional transactivation domain is required for activating p53-inducible genes. The N-terminus contains two complementary transcriptional activation domains, with a major one at residues 1–42 and a minor one at residues 55–75, specifically involved in the regulation of several pro-apoptotic genes (Venot *et al.*, 1998). Proline rich domain important for the apoptotic activity of p53: residues 64-92. The central DNA-binding domain facilitates sequence-specific binding of p53 to p53-response elements in DNA. The domain contains one zinc atom and



several arginine amino acids: residues 102-292. This region is responsible for binding the *p53* co-repressor LMO3 (Larsen *et al.*, 2010). The tetramerization domain facilitates the interaction of *p53* monomers to form dimers, and the interactions of dimers to form tetramers. Tetramerization is essential for the ability of *p53* to positively regulate gene expression.

Along with the function the proper regulation of *p53* abundance are necessary for the initiation of an accurate *p53* response. The levels of *p53* expression are key to its activity and are tightly controlled in the cell, mainly by covalent modifications (Lazar *et al.*, 1993). In unstressed cells, a constant proteasomal degradation of *p53* keeps low the expression level of *p53* (Figure 2). The mark for proteasomal degradation (i.e. polyubiquitylation) is attached to C-terminal lysine residues of *p53* by the E3 ligase MDM2 (Toledo and Wahl, 2006). Transfer of the ubiquitin moiety from an upstream E2 ligase depends on the RING (really interesting new gene) domain of MDM2 (Itahana *et al.*, 2007). Interestingly, although MDM2 is necessary and sufficient for ubiquitylation of *p53*, its homologue MDMX is required for efficient *p53* regulation. MDMX also contains a RING (really interesting new gene) domain. However, it does not itself show significant ubiquitin ligase activity towards *p53* (Linares *et al.*, 2003). Nevertheless, MDMX forms heteromers with MDM2 and enhances MDM2-mediated ubiquitylation of *p53* (Linares *et al.*, 2003). The importance of both MDM2 and MDMX is highlighted by the fact that single knockout of each gene confers embryonic lethality (Toledo and Wahl, 2006). MDM2 and also MDMX are frequently over-expressed in tumours that possess wt-*p53*. The abundance, subcellular localization and activity of *p53* are regulated by post-translational modifications such as phosphorylation, acetylation, ubiquitylation and

attachment of ubiquitin-like moieties such as SUMO and Nedd8 on certain residues mainly in the N- or C-terminal part of *p53* (Toledo and Wahl, 2006). Phosphorylation primarily occurs at N-terminal Ser and Thr residues and affects *p53* stability as well as the decision of whether *p53* induces apoptosis or senescence. The phosphorylation at N-terminal residues influences binding of MDM2 and thereby modulates ubiquitylation of *p53* at C-terminal lysine residues (Toledo and Wahl, 2006). Acetylation of these C-terminal residues also prevents ubiquitylation (Bode and Dong, 2004). MDM2 binds to *TP53* mRNA, controlling the rate of translation (Mcevoy *et al.*, 2012), and MDM2 regulates the levels of itself, MDM4 (also known as MDMX) and *p53* (Mcevoy *et al.*, 2012; Manfredi, 2010).

The pivotal role of MDM2 and MDM4 in the control of *p53* function argues that polymorphisms at these loci should be scrutinized for potential modulation of *p53* function. Polymorphisms at loci that alter the activity of any single upstream event that activates *p53* should not entirely abrogate the *p53* response, owing to the high level of redundancy in stress responses, but cellular responses could be attenuated by altering one or more of the triggers for *p53* activation (Catherine *et al.*, 2009).

The range of post-translational modifications and *p53* upregulation elicited by stress are well-studied features of the *p53* network. Less is known about the physiological roles of *p53* that are not necessarily linked to tumour suppression and that can be executed by low levels of *p53* and in the absence of a severe insult or stress.

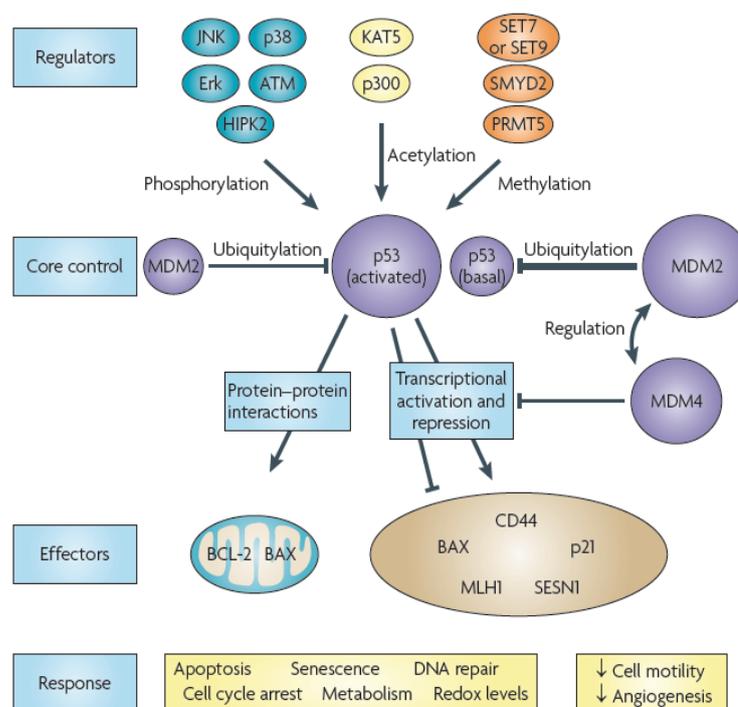


Figure 2. The p53 pathway (Catherine *et al.*, 2009). The p53 pathway is complex. At least 50 different enzymes can covalently modify p53 to alter its stability, cellular location or activity¹⁷. Under normal cellular conditions, MDM2 represses p53 by binding and sequestering p53, and by ubiquitylating p53, targeting it for degradation. DNA damage, oxidative stress and oncogene activation are among the processes that can activate p53 by a range of regulators. Basal levels of p53 and p53 in cells that are undergoing low levels of stress can also affect cell physiology. Under high levels of stress, the interactions between MDM2, MDM4 and p53 are disrupted by post-translational modifications of these proteins. This allows activated p53 to act as a transcription factor, activating or repressing genes involved in apoptosis, cell cycle arrest and senescence. p53 can also move to the mitochondria, where it physically interacts with members of the Bcl-2 family to form pores in the mitochondrial membrane, leading to the release of cytochrome *c* and subsequent apoptosis. Some of the more intensively studied activators, regulators and effectors of p53 are shown in the figure. ATM, ataxia-telangiectasia mutated; BAX, BCL-2-associated X; HIPK2, homeodomain-interacting protein kinase 2; JNK, Jun N-terminal kinase; KAT5, K (lysine) acetyltransferase 5; MLH1, MutL protein homologue PRMT5,

Protein arginine methyltransferase 5; SESN1, sestrin 1; SMYD2, SET and MYND domain-containing 2. (Catherine *et al.*, 2009)

1.14 Polymorphism of TP53 Gene

Mutation of *TP53* occurs in about 50% of human tumours, and most of these mutations are located in the DBD (Bode and Dong, 2004), influencing the specific binding of *TP53* to target sequences. The most common mutations concentrate on only a few nucleotides. Several of these hot spot mutations result in the exchange of amino acids that make contact with the DNA. Therefore, exchange of these amino acids weakens binding of p53 to DNA or alters sequence specificity of the mutant *TP53*. Altered sequence specificity in turn can even result in mutant *TP53* with oncogenic functions, thus turning the tumour suppressor to an oncogene. Mutations that cause structural destabilization of the DBD may result not only in its inactivation but also in deregulation of p53 by interfering with protein-protein interaction. Interestingly, mutations that interfere with p53 oligomerization as well as deletion of p53 are not common in human cancers. It is assumed that, once an inactivating mutation in one *TP53* allele has occurred, the resulting protein efficiently blocks the activity of the wt-p53 expressed from the second allele (Essmann, and Schulze, 2012). Thus, diverse mechanisms exist that impede a proper *TP53* response, and hence, also diverse strategies have been developed to overcome them.

Mutations in tumor suppressor *TP53* gene have been associated with inherited cancer susceptibility (Xifeng *et al.*, 2002). Thirteen different polymorphisms have been detected in human *TP53* gene. Among them, five were found in exons and eight in introns. The five coding region polymorphisms include a C→T transition in codon 21 (Ahuja *et al.*, 1990), a G→A transition in codon

36 (Felix *et al.*, 1999), a C→T transition in codon 47 (Felley *et al.*, 1993), a G→C transversion in codon 72 (Weston *et al.*, 1992) and an A→G transition in codon (Carbone *et al.*, 1991). However, among the five polymorphisms, those in codons 21, 36 and 213 are lack of significance- that means, they do not result in a change in the encoded amino acid sequence. The codon 47 polymorphism results in a proline to serine substitution and the polymorphism at codon 72 results in an arginine to proline substitution, which is in exon 4 (Xifeng *et al.*, 2002). Whereas, the codon 72 polymorphism is common (Carbone *et al.*, 1991), the polymorphism in codon 47 is rare (Felley *et al.*, 1993).

More than 90% of the polymorphisms in *TP53* gene occur are noncoding region polymorphism. A 16 base pair insertion in intron 3 is the well-characterized intronic *TP53* polymorphism (Lazaret *et al.*, 1993). This intronic polymorphism has been associated with an increase in the risk of several types of cancer. Several studies (Birgander *et al.*, 1995; Schneider *et al.*, 2004 and Walerych *et al.*, 2012) have estimated the haplotype frequencies for the polymorphisms in exon 4 and introns 3 and 6 and determined that the *TP53* haplotypes were associated with risk for lung (Birgander *et al.*, 1995), colorectal (Schneider *et al.*, 2004), and breast cancers (Walerych *et al.*, 2012). and Walerych *et al.*, 2004) found a statistically significant difference in the distribution of the intron 6 polymorphism between healthy Caucasian control subjects and patients with ovarian cancer but found no difference between control subjects and patients with breast cancer.

Polymorphism at codon 47 (P47S) is rare located in the N-terminal transactivation domain of p53. This polymorphism results from a C>T base substitution at position 1 of codon 47. It has only been reported in populations of African origin. In which it is found at an allele frequency of approximately 5% (Weston and Godbold, 1997). Dai and Gu (2010) showed that phosphorylation of the N-terminal domain of p53 regulate its transactivation properties. S46 is phosphorylated by p38 and homeodomain-interacting protein kinase 2 (HIPK2), which enhances the transcription of apoptosis-related genes and hence promotes p53-mediated apoptosis (Kato *et al.*, 2003). These two kinases are directed to phosphorylation sites by a proline residue adjacent to S46. For this reason, replacement of P47, as occurs with the P47S polymorphism, would be expected to decrease phosphorylation at S46 result in decrease transactivation of pro-apoptotic target genes and thus potentially increase cancer risk (Felley *et al.*, 1993). Codon 217 polymorphism (V217M) (resulting from a G>A transition) is the only validated coding polymorphism located in the DNA binding domain (DBD) of p53. Thus, it could dramatically affect the activity of p53. Functional studies have been limited to transactivation assays

in yeast, which indicate that this polymorphism results in little loss of activity (DAI and GU 2010). The genes that show the most variation in activation are *CDKN1A*, *BAX* and *PMAIP1* (also known as NoXA). However, the p53-M217 variant leads to greater expression of these genes than the more common p53-V217 variant. Extrapolating from this result, one can speculate that the V217M polymorphism might be protective against cancer.

Codon 360 polymorphism (G360A) is located in the linker region adjacent to the tetramerization domain of p53. Again, the functional data for this polymorphic variant have been provided by transactivation studies in yeast, which showed a slight decrease in the transactivation of *BAX*, *MDM2* and *P53AIP*, and a more marked decrease in stratifin (*SFN*, also known as 14-3-3 sigma) and *GADD45* (growth arrest and DNA damage-inducible 45) (Kato *et al.*, 2003).

Codon 72 polymorphism (R72P) is in exon 4 located in the segment of p53 gene that encodes the polyproline domain, which lies between the N-terminal transactivating domain and DNA binding domain (DBD). Most tumour associated mutations are found in this region (Venot *et al.*, 1998). The polyproline domain is less well conserved across the species than the DBD. Deletion studies in cell and mice showed that the polyproline domain is essential for p53 to show a full apoptotic response to stress and inhibit tumorigenesis (Rot *et al.*, 2000 and Toledo *et al.*, 2007).

From comparative sequence analyses in non-human primates, it was found that p53-P72 is the ancestral form, although p53-R72 occurs at a high frequency (>50%) in some populations (Puente *et al.*, 2006). A latitude gradient in variant frequency (an increasing frequency of the p53-72 variant towards the equator (Beckman, 1994 ; Ozbey *et al.*, 2011 ; Ricksi *et al.*, 2010; Shi *et al.*, 2009) invited early speculation that p53-P72 might protect against adverse consequences of sunlight or other environmental cancer risk factors. Interestingly, p53 influences the tanning response to sunlight by inducing the expression of pro-opiomelanocortin (POMC) (NAN *et al.*, 2008), which leads to an idea that p53 polymorphisms influence *POMC* transactivation. The contribution of p53-mediated control of leukaemia-inhibitory factor expression, which is crucial for blastocyst implantation, has recently been considered in the discussion of evolutionary selection of specific alleles in the p53 pathway (Feng *et al.*, 2011; Hu 2009; Mathieu *et al.*, 2012).

1.15 Codon 72 polymorphism and lung cancer

p53 protein exists in two polymorphic forms, p53-Pro (p53-P72) or p53-Arg (p53-R72) in the general population due to single nucleotides change at codon 72 of exon 4 of the *TP53* gene (MATLASHEWSKI *et al.* 1987) shows different structure and functional properties. The current

consensus is that p53-R72 is more effective at inducing apoptosis and protecting stressed cells from neoplastic development than p53-P72. However, to understand how universal these functional differences between p53-P72 and p53-R72 might be different cell type or whether they are relevant *in vivo*, the experimental data is insufficient (CHEN *et al.* 2011). NIH genetic association database, which is not comprehensive, has records on over 230 studies evaluating the effect of the codon 72 polymorphism on susceptibility to a wide variety of cancers (MATLASHEWSKI *et al.* 1987). Many of these studies have reported 'statistically significant' associations. The association of p53 codon 72 polymorphism with lung cancer risk has been studied by several groups, although with inconsistent result. Weston found the pro allele to be in excess in patients with adenocarcinoma in USA, but this could not be confirmed in its follow-up study (WESTON *et al.* 1994). In Japan, a study showed a significant association of the Pro allele with squamous cell carcinoma rather than lung adenocarcinoma (KAWAJIRI *et al.* 1993). Ying chuan Hu showed that the p53 Pro allele is associated with an increased frequency of p53 mutation in non-small cell lung cancer. They also stated that the codon 72 polymorphism did not influence patient's survival in either the entire patient group or among patients with p53 mutant tumor (Ying chuan Hu *et al.* 2005). Weston found that the frequency of polymorphic variants was similar in lung cancer cases and controls after adjustment for race (African-Americans and Caucasians in U.S. population). However, the proline variant at codon 72 was in excess in adenocarcinoma among lung cancer patient (AINSLEY WESTON *et al.* 1992). In Taiwan, a study showed that the Pro allele of the p53 codon 72 polymorphism increased the risk of lung cancer (especially adenocarcinoma) among female Taiwanese. Patients with lung cancer diagnosed at the early stage had an increased frequency of Pro/Pro genotype. Patients with the Pro/Pro genotype tended to have poorer prognosis than those with the Arg/Pro genotype (Ying chuan Hu *et al.* 1999). In India, Neeraj Jain found a strong association of p53 Arg homozygotes with advanced lung cancer. They also expressed that the presence of Arg/Pro heterozygotes may associate with early progression of the disease possibly due to additional genetic alteration.

As to the correlation of p53 codon 72 polymorphism with smoking, there are conflicting results. Smoking is considered to be one of the principal causes of lung cancer. Murata found that lung cancer patients who did not smoke included a significantly larger proportion of Arg/Arg homozygotes and smaller proportion of Arg/Pro heterozygotes compared with the healthy controls (Murata *et al.* 1996). However, Jin reported that increased risks associated with the Pro/Pro

genotype were noted in lighter smokers (Jin *et al.* 1995).

1.16 Reasons for studying p53 for Cancer Diagnostics and Prognosis

Many researchers focused on the study of p53 structure and function for cancer diagnosis and prognosis. Mutations in the tumour suppressor TP53 gene are one of the most common genetic alterations, which present at high frequency in human cancers (vogelstein *et al.*, 2000; Hofseth, *et al.*, 2004; Levine and Oren, 2009). More than 26,000 somatic mutation data of p53 appear in the international agency for research on cancer (IARC) TP53 database version R14 (current version (November, 2009) (olivier *et al.*, 2010). Up to 50% of all human cancers contain mutations in both alleles of the TP53 gene (Edlund *et al.*, 2012; Harris *et al.*, 2012). Unlike most other tumor suppressor genes, such as RB, APC, and BRCA1 genes, they are inactivated by frameshift or nonsense mutations, which leads to disappearance or aberrant synthesis of the gene product, nearly 80% of TP53 gene mutations are missense mutations (Essmann and Schulze, 2012). Other mutations include frameshift insertions and deletions (9%), nonsense mutations (7%), silent mutations (5%), and other infrequent alterations (Cui *et al.*, 2007). The frequency of TP53 mutation varies from ~10% (hematopoietic malignancies) to 50–70% (ovarian, colorectal, and head and neck malignancies) (Rotter *et al.*, 2009). Germline mutation of TP53 causes Li-Fraumeni syndrome, which is a familial cancer syndrome including breast cancer, soft tissue sarcoma, and various other types of cancer (Malkin *et al.*, 1990; Manfredi *et al.*, 2006). Most TP53 mutations in human cancers result in mutations within the DNA binding domain, thus preventing p53 from transcribing its target genes. However, mutant p53 has not only led to a loss of normal function of the wild-type protein but also led to new abilities to promote cancer (Rotter *et al.*, 2009).

The key role of p53 as a tumor suppressor is to block cell cycle progression and/or to induce apoptosis, in response to cellular stresses such as DNA damage. Numerous reports have described the mechanism by which p53 induces apoptosis. As p53 functions mainly as a transcription factor, it is important to explore the genes regulated by p53 that contribute to the regulation of apoptosis. Early studies showed that wild-type p53 can bind the *bax* gene promoter region and regulate *bax* gene transcription (Reed *et al.*, 1995; Miyashita *et al.*, 1994). *bax* is a member of the Bcl-2 family, which forms heterodimers with Bcl-2, inhibiting its activity (Oltvai *et al.*, 1993). The Bcl-2 protein family plays an important role in apoptosis and cancer (Plati *et al.*, 2011). For example, Bcl-2 controls the release of cytochrome c from the mitochondria, which activates the apoptotic pathway by activating caspase 9.

The p53 protein suppresses tumor formation not only by inducing apoptosis but also by causing cell cycle arrest. Depending on the type of cellular stress, p53 can induce G1 arrest through activation of transcription of the cyclin-dependent kinase inhibitor p21 (Chung and Bunz, 2010). p53 also regulates the G2/M transition. For example, p53 can block cell entry into mitosis by inhibition of Cdc2. Cdc2 needs to bind to cyclin B1 in order to function. Repression of cyclin B1 by p53 also arrests of cells in G2 (Chung and Bunz, 2010).

This is because most, but not all, human cancers harbor altered p53; the concept of restoration of p53 for cancer therapy is very attractive. An animal model showed the reactivation of wild type p53 to result in efficient tumor regression, including regression of lymphoma (WANG *et al.*, 2011) and liver carcinoma (Chien *et al.*, 2011). p53 has been reported to induce apoptosis independent of its transcription of genes (Dixit *et al.*, 2012). Surprisingly, activated p53 can induce apoptosis in the cytoplasm by a *bax*-dependent mechanism (Stegh, 2012). Recent reports have showed that p53 regulates the process of self-renewal of neural stem cells (Hede *et al.*, 2011) and hematopoietic stem cells (Liu *et al.*, 2009).

It was reported that somatic TP53 missense mutations were found in about 50% of human cancers, and inactivating mutations in the TP53 gene were the most common genetic events in human cancers affecting a specific gene, with the vast majority arising from a single-point mutation in the DNA-binding domain encoding segment of TP53 (Harris *et al.*, 2012; Leroy *et al.*, 2013). The mutant TP53 protein rendered by the inactivating mutations unable to carry out its normal functions, that is, transcriptional transactivation of downstream target genes that regulate cell cycle and apoptosis (Hamzehloie *et al.*, 2012). Several recent studies indicated that in addition to abrogating the tumor suppressor functions of wild-type TP53, the common types of cancer-associated TP53 mutations also furnish the mutant protein with new activities, so-called "gain-of-function" (GOF) activities. These new activities can contribute actively to various stages of tumor progression, including distant metastases, and to increased resistance to anticancer treatments. GOF activities of mutant TP53 are exerted by aberrant protein interaction or gene regulation, such as MAPKK3, inhibitor of DNA-binding 4 (ID4), polo-like kinase 2 (Plk2), promyelocytic leukemia protein (PML), and prolyl isomerase Pin1 (Hamzehloie *et al.*, 2012; Kim *et al.*, 2011). Although the occurrence of TP53 mutations is not limited to a few particular sequences or codons along this gene, most mutations cluster in the TP53 DNA binding domain, which encompasses exons five through eight and spans approximately 180 codons or 540 nucleotides (Pfeifer and Besaratinia, 2009). Most TP53 missense mutations lead to the synthesis of a stable protein, which loses its

specific DNA-binding and transactivation function and accumulates in the nucleus of cells. These mutant accumulated proteins play a vital role and are retained in distant metastasis. In moreover, the most frequent mutants have been shown to be capable of cooperating with oncogenes for cellular transformation (Markowitz *et al.*, 1994). About 30% of TP53 missense mutations found in cancer correspond to nucleotide substitutions at highly mutable CpG dinucleotides and at codons encoding residues that play essential structural and chemical roles in the contact between the TP53 protein and specific DNA sequences that constitute the TP53 response elements (Olivier *et al.*, 2010). These mutations result in a significant loss of DNA-binding activity and transactivation capacity (Hamzehloie *et al.*, 2012).

Loss of heterozygosity (LOH) frequently detected in lung cancer cell lines and tumor samples at the location of the TP53 gene on chromosome 17p13, which suggested that this gene was likely to be involved in the pathogenesis of lung cancer, and genetic abnormality of the TP53 in lung cancers has been shown to be associated with a poorer survival prognosis and increased cellular resistance to therapy (Liu *et al.*, 2011). The highest frequency of TP53 alterations is found in small cell lung cancer specimens (Lohmann *et al.*, 1993; Stewart, 2010). In contrast, the frequency of TP53 mutations is the highest in squamous cell carcinomas and lowers in adenocarcinomas among non-small cell carcinoma samples (Lohmann *et al.*, 1993 and Johnson *et al.*, 2012). It has been reported that somatic mutations and increased expression of TP53 were frequently found in 23% and 65% of non-small cell carcinoma respectively, (Baldi *et al.*, 2011; Bornachea *et al.*, 2012). Moreover, TP53 mutations are found in tumors both with and without allele loss at 17p13, which are mostly located within the DNA-binding domain of TP53 (Hong *et al.*, 2008). Because of coding mutations of TP53 occur relatively early in the development of lung cancer and are potentially required for maintaining the malignant phenotype, the acquired TP53 mutations are preserved during tumor progression and metastatic spread (D'Agostini *et al.*, 2008; Newnham *et al.*, 2011). It has been reported that the incidence of TP53 mutations in primary tumors and metastatic lymph nodes was 23.2% and 21.4%, respectively. The TP53 gene status in primary tumors and metastatic lymph nodes showed 92.9% harmony among 56 patients with non-small lung cancer who had undergone surgical resection, which explained the fact that TP53 mutations usually precede lymph node metastasis (Rivlin *et al.*, 2011). Most TP53 mutations occur before the tumor metastasizes. They are then preserved through subsequent stages of tumor development; as a result, no selection against TP53 mutations occurs during metastasis.

2. Discussion and Conclusion

The population-base case-control study was carried out in Bangladesh population to examine the prevalence of p53 codon 72 polymorphism in lung cancer patients and to make relation of the polymorphisms with different demographic characteristics. We have found over half of the cancer patients (68.40%) have squamous cell carcinoma, which is slightly more than frequency (53%) estimated in the Cancer Registry Report (CRR) of the NICRH, 2005-2007 in Bangladesh (CRR, 2009). In an opposite fashion the frequency of adenocarcinoma in the present study is less than that of Cancer Registry Report (CRR, 2009) of the NICRH. The remarkable that there is no large cell carcinoma in lung cancer patients in Bangladesh. This outcome is similar to the Neeraj (52.50% of squamous cell carcinoma case and 2.50% of large cell lung cancer case) find in India. However, in Greek, 56.50% of lung cancer type was adenocarcinoma which is different from our study (PAPADAKIS et al. 2007). Nearly 95% of the lung cancer patients are more than 40 years old. This indicates, with the increase of age, the risk of lung cancer development increases. From our patient study, it is found that about half of the patients are in the age group of 51-60. That means lung cancer develop in the older people. This finding is supported by the CRR of the NICRH, Bangladesh. They showed that the highest number of lung cancer patients were 55-65 years old. According to present study and Cancer Registry Report (CRR) of the NICRH male are prone to lung cancer, which is supported by the WHO (WHO, 2008).

The relation of p53 codon 72 polymorphism with lung cancer is inconsistent as well as conflicting worldwide. Some studies found no correlation between p53 codon 72 polymorphism and lung cancer (BIRGANDER R et al. 1995; WESTON and GODBOLD et al. 1997). On the other hand the worldwide scenario in the frequency of p53 polymorphism in lung cancer also indicate that Arg/Arg and Arg/Pro genotypes are more susceptible to lung cancer in Asian (JAIN N et al. 2007; TAGAWA M et al. 1998), European (PAPADAKIS et al. 2002), and North American population (JIN X et al. 1995; MILLER DP et al. 2002). Similarly, in India, Neeraj Jain and Pandima Devi found association between Arg/Arg genotype with lung cancer (Neeraj Jain et al. 2005; Pandima Devi et al. 2010)

Other authors reported an increased frequency of Pro/Pro homozygotes in lung cancer (KAWAJIRI ET AL. 1993; WANG YC et al. 1999). Studies on the people of Asian and Mexican-Americans suggested at least at fivefold increase in lung

cancer risk for the Pro/Pro homozygotes (KAWAJIRI ET AL. 1993; WANG YC et al. 1999). Also, an increased risk was observed for African-American (FAN et al. 2000) and white population with Pro/Pro genotypes (WANG YC et al. 1999). In agreement with the above results, we found increased frequency of Pro/Pro genotype in lung cancer patients compare to control and which was statistically more significant than Arg/Arg genotype. Our result also supported by majority of studies that associate p53 Pro/Pro genotype with lung cancer have been reported from Southeast Asia (China) (W. XUE et al. 2007), Taiwan or Japan (YI-CHING WANG et al. 1999). The discrepancies in the result may be attributed to various reasons such as selection of patients and control subjects, variation in laboratory protocols, and geographic and ethnic background. Fan, who demonstrated the association of p53 proline homozygosity with lung cancer, had studied only primary lung cancer stage I and II patients and excluded patients of advanced stage, (FAN et al. 2000) whereas Papadakis and Neeraj Jain studied advanced lung cancer patients (stage III and IV) and suggested association of p53 arginine homozygosity with lung cancer. Interestingly, we found increased frequency of both Arg/Arg and Pro/Pro homozygous genotype in lung cancer patient compare to control subject; although the results were not statistically significant (Pro/Pro genotype is more statistically significant) (PAPADAKIS et al. 2002; NEERAJ JAIN et al. 2005)

The p53 plays important roles in apoptosis, cell cycle control and DNA repair, and these critical functions differ among p53 protein encoded by the Arg and Pro alleles. In the presences of Arg allele, conformational p53 mutants have been more potent in binding to p53 and neutralizing p53-induced apoptosis, which enhances tumorigenesis and provides a selective growth advantage to tumour cells (PANDIMA et al. 2010). Earlier studies reveal that people with Arg(Arg/Pro or Arg/Arg phenotype) in codon 72 of p53 gene are more susceptible for lung cancer at the later stage of life and longtime of smoking habit (PAPADAKIS et al. 2002; NEERAJ JAIN et al. 2005). The increased frequency Arg/Arg genotype was present in the patients who are more than 50 years old and smoke for a long time.

Acknowledgements

The authors are thankful to all of the teachers in the Department of Biotechnology and Genetic Engineering, Mawlana Bhashani Science and Technology University, Tangail-1902, Bangladesh, for their valuable cooperation during this article proceeding.

References

- AHUJA HG, T. M., CLINE MJ, 1990 Variation in the protein coding region of the human p53 gene. *Oncogene* 5: 1409-1410.
- AINSLEY WESTON, L. S. P., KATHLEEN FORRESTER, ROBERT N. HOOVER, BENJAMIN F. TRUMP, CURTIS C. HARRIS, AND NEIL E. CAPORASO, 1992 Allelic Frequency of a p53 Polymorphism in Human Lung Cancer. *Cancer Epidemiol Biomarkers Prev* 1: 481- 483.
- ALBERG, A. J., J. G. FORD and J. M. SAMET, 2007 *Epidemiology of lung cancer: ACCP evidence-based clinical practice guidelines* (2nd edition). *Chest* 132: 29S-55S.
- ALBERG AJ, S. J. I., 2010 *Murray &Nadel's Textbook of Respiratory Medicine (5th ed.)* Saunders Elsevier. ISBN 978-1-4160-4710-0.
- ARAMAYO, R., M. B. SHERMAN, K. BROWNLESS, R. LURZ, A. L. OKOROKOV *et al.*, 2011 Quaternary structure of the specific p53-DNA complex reveals the mechanism of p53 mutant dominance. *Nucleic acids research* 39: 8960-8971.
- BALDI, A., A. DE LUCA, V. ESPOSITO, M. CAMPIONI, E. P. SPUGNINI *et al.*, 2011 Tumor suppressors and cell-cycle proteins in lung cancer. *Pathology research international* 2011: 605042.
- BECKMAN, G. E. A., 1994 Is p53 polymorphism maintained by natural selection? *Hum. Hered.* 44: 266–270.
- BEHZADI, A., Y. UNG, V. LOWE and C. DESCHAMPS, 2009 The role of positron emission tomography in the management of non-small cell lung cancer. *Canadian journal of surgery. Journal canadien de chirurgie* 52: 235-242.
- BIDERMAN, L., J. L. MANLEY and C. PRIVES, 2012 Mdm2 and MdmX as Regulators of Gene Expression. *Genes & cancer* 3: 264-273.
- BIESALSKI, H. K., B. BUENO DE MESQUITA, A. CHESSON, F. CHYTIL, R. GRIMBLE *et al.*, 1998 European Consensus Statement on Lung Cancer: risk factors and prevention. *Lung Cancer Panel. CA: a cancer journal for clinicians* 48: 167-176; discussion 164-166.
- BIRGANDER R, S. A., RANNUG A, ALEXANDRIE AK, SUNDBERG MI, SEIDEGARD J, ET AL, 1995 P53 polymorphisms and haplotypes in lung cancer. *Carcinogenesis* 16: 2233–2236.
- BIRGANDER R, S. A., RANNUG A, ALEXANDRIE AK, SUNDBERG MI, SEIDEGARD J, ET AL. , 1995b P53 polymorphisms and haplotypes in lung cancer. *Carcinogenesis (Lond.)* 16: 2233–2236.
- BODE, A., and Z. DONG, 2004 Post-translational modification of p53 in tumorigenesis. *Nat Rev Cancer* 4: 793–805
- BORNACHEA, O., M. SANTOS, A. B. MARTINEZ-CRUZ, R. GARCIA-ESCUADERO, M. DUENAS *et al.*, 2012 EMT and induction of miR-21 mediate metastasis development in Trp53-deficient tumours. *Scientific reports* 2: 434.
- BOUCHET, B. P., C. C. D. F., A. PUISIEUX, AND C. M. GALMARINI, 2006 p53 as a target for anti-cancer drug development. *Critical Reviews in Oncology/Hematology* 58: 190–207.
- CARBONE D, C. I., MITSUDOMI T, 1991 Polymorphism at codon 213 within the p53 gene. *Oncogene* 6: 1691-1692.
- CATHERINE WHIBLEY, P. D. P. P. A. M. H., 2009 p53 polymorphisms: cancer implications. *Nature Reviews | Cancer* 9: 95-107.
- CHEN, H., M. S. GOLDBERG and P. J. VILLENEUVE, 2008 A systematic review of the relation between long-term exposure to ambient air pollution and chronic diseases. *Reviews on environmental health* 23: 243-297.
- CHEN, X., F. LIU, B. LI, Y. G. WEI, L. N. YAN *et al.*, 2011 p53 codon 72 polymorphism and liver cancer susceptibility: a meta-analysis of epidemiologic studies. *World journal of gastroenterology : WJG* 17: 1211-1218.
- CHIEN, Y., C. SCUOPPO, X. WANG, X. FANG, B. BALGLEY *et al.*, 2011 Control of the senescence-associated secretory phenotype by NF-kappaB promotes senescence and enhances chemosensitivity. *Genes & development* 25: 2125-2136.
- CHUNG, J. H., and F. BUNZ, 2010 Cdk2 is required for p53-independent G2/M checkpoint control. *PLoS genetics* 6: e1000863.
- CUI, R. E. A., 2007 Central role of p53 in the suntanresponse and pathologic hyperpigmentation. *Cell cycle* 128: 853–864.

- D'AGOSTINI, F., R. BALANSKY, V. E. STEELE, G. GANCHEV, C. PESCE *et al.*, 2008 Preneoplastic and neoplastic lesions in the lung, liver and urinary tract of mice exposed to environmental cigarette smoke and UV light since birth. *International journal of cancer. Journal international du cancer* 123: 2497-2502.
- D. MALKIN, F. P. L., L. C. STRONG ET AL, 1990 Germ line p53 mutations in a familial syndrome of breast cancer, sarcomas, and other neoplasms. *Science* 250: 1233– 1238.
- DAI, C., and W. GU, 2010 p53 post-translational modification: deregulated in tumorigenesis. *Trends in molecular medicine* 16: 528-536.
- DAVIES, R. L. Y. (Editor), 2010 *Oxford Textbook Medicine*. OUP Oxford.
- DAYEM UDDIN, 2002 Non-Small Cell Lung Cancer (NSCLC): An Update. *The Journal of Teachers Association RMC, Rajshahi* 15: 115-118.
- DIXIT, D., V. SHARMA, S. GHOSH, V. S. MEHTA and E. SEN, 2012 Inhibition of Casein kinase-2 induces p53-dependent cell cycle arrest and sensitizes glioblastoma cells to tumor necrosis factor (TNF α)-induced apoptosis through SIRT1 inhibition. *Cell death & disease* 3: e271.
- EDLUND, K., O. LARSSON, A. AMEUR, I. BUNIKIS, U. GYLLENSTEN *et al.*, 2012 Data-driven unbiased curation of the TP53 tumor suppressor gene mutation database and validation by ultradeep sequencing of human tumors. *Proceedings of the National Academy of Sciences of the United States of America* 109: 9551-9556.
- ESSMANN., F., and K. SCHULZE-OSTHOFF., 2012 Translational approaches targeting the p53 pathway for anti-cancer therapy. *British Journal of Pharmacology* **165**: 328–344.
- EZZATI, M., and A. D. LOPEZ, 2004 Regional, disease specific patterns of smoking-attributable mortality in 2000. *Tobacco control* 13: 388-395.
- FAN, R., M. T. WU and D. MILLER, 2000 The p53 codon 72 polymorphism and lung cancer risk. *Cancer Epidemiol Biomarkers Prev* 9: 1037–1042.
- FELIX CA, B. D., MITSUDOMI T, IKAGAKI N, WONG A, WASSERMAN R, 1999 Polymorphism at codon 36 of the p53 gene. *Br J Cancer* 81: 179–183.
- FELLEY-BOSCO, E., A. WESTON, H. M. CAWLEY, W. P. BENNETT and C. C. HARRIS, 1993 Functional studies of a germ-line polymorphism at codon 47 within the p53 gene. *American journal of human genetics* 53: 752-759.
- FENG, Z., C. ZHANG, H. J. KANG, Y. SUN, H. WANG *et al.*, 2011 Regulation of female reproduction by p53 and its family members. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 25: 2245-2255.
- FERLAY J, S. H., BRAY F, FORMAN D, MATHERS C, PARKIN DM, 2008 Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 10 [Internet] Lyon. GLOBOCAN 1.2.
- HAMZEHLOIE, T., M. MOJARRAD, M. HASANZADEH NAZARABADI and S. SHEKOUHI, 2012 The role of tumor protein 53 mutations in common human cancers and targeting the murine double minute 2-p53 interaction for cancer therapy. *Iranian journal of medical sciences* 37: 3-8.
- HARRIS, A. I. R. A. C. C., 2012 Clinical outcomes and correlates of TP53 mutations and cancer. *Cold Spring Harbor perspectives in biology* 2.
- HECHT, S. S., 2003 Tobacco carcinogens, their biomarkers and tobacco-induced cancer. *Nature reviews. Cancer* 3: 733-744.
- HEDE, S. M., I. NAZARENKO, M. NISTER and M. S. LINDSTROM, 2011 Novel Perspectives on p53 Function in Neural Stem Cells and Brain Tumors. *Journal of oncology* 2011: 852970.
- HOFSETH, L. J., S. P. H., AND C. C. HARRIS, 2004 p53: 25 Years after its discovery. *Trends in Pharmacological Sciences* 25: 177–181.
- HOGAN, D. B., 1999 Did Osler suffer from "paranoia antitherapeuticum baltimorensis"? A comparative content analysis of *The Principles and Practice of Medicine* and *Harrison's Principles of Internal Medicine*, 11th edition. *CMAJ : Canadian Medical Association journal = journal de l'Association medicale canadienne* 161: 842-845.
- HONG, H. H., T. V. TON, Y. KIM, N. WAKAMATSU, N. P. CLAYTON *et al.*, 2008 Genetic alterations in K-ras and p53 cancer genes in lung neoplasms from B6C3F1 mice exposed to cumene. *Toxicologic pathology* 36: 720-726.

- HU, W., 2009 The role of p53 gene family in reproduction. *Cold Spring Harbor perspectives in biology* 1: a001073.
- HU, Y., M. MP. and A. SA., 2005 The p53 codon 72 proline allele is associated with p53 gene mutations in non-small cell lung cancer. *Clin Cancer Res* 11: 2502—2509
- HUBAUX, R., D. D. BECKER-SANTOS, K. S. ENFIELD, S. LAM, W. L. LAM *et al.*, 2012 Arsenic, asbestos and radon: emerging players in lung tumorigenesis. *Environmental health : a global access science source* 11: 89.
- CHEN, X., F. LIU, B. LI, Y. G. WEI, L. N. YAN *et al.*, 2011 p53 codon 72 polymorphism and liver cancer susceptibility: a meta-analysis of epidemiology studies. *World journal of gastroenterology: WJG* 17:1211-1218
- ITAHANA, K., H. MAO, JIN A, ITAHANA Y and C. HV., 2007 Targeted inactivation of Mdm2 RING finger E3 ubiquitin ligase activity in the mouse reveals mechanistic insights into p53 regulation. *Cancer Cell* **12**: 355–366.
- JAIN N, L. Y., ZHANG L, MENENI SR, CHO BP, 2007 Probing the sequence effects on NarI-induced -2 frame shift mutagenesis by dynamic 19F NMR, UV, and CD spectroscopy. *Biochemistry research international* 46: 13310-13321.
- JEMAL A, T. A., MURRAY T, THUN M, 2002 Cancer statistics, 2002. *CA Cancer J Clin* 52: 23 – 47.
- JIN, X., WU, X., ROTH, J. A., AMOS, C. I., KING, T. M., BRANCH, C., HONN, S. E., AND SPITZ, R., 1995 Higher lung cancer risk for younger African- Americans with the Pro/Pro p53 genotype. *Carcinogenesis (Lond.)* 16: 2205–2208.
- JIN X, W. X., ROTH JA, ET AL., 1995 Higher lung cancer risk for younger African-Americans with the Pro/Pro p53 genotype. *Carcinogenesis (Lond.)* 16: 2205–2208.
- JOHNSON, J. L., S. PILLAI and S. P. CHELLAPPAN, 2012 Genetic and biochemical alterations in non-small cell lung cancer. *Biochemistry research international* 2012: 940405.
- KATO, S., S. Y. HAN, W. LIU, K. OTSUKA, H. SHIBATA *et al.*, 2003 Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. *Proceedings of the National Academy of Sciences of the United States of America* 100: 8424-8429.
- KAWAJIRI, K., NAKACHI, K., IMAI, K., WATANABE, J., AND HAYASHI, S-I. , 1993 Germ line polymorphisms of p53 and CYP1A1 genes involved in human lung cancer. *Carcinogenesis (Lond.)* 14: 1085–1089.
- KERN JA, M. G. P., 2008 *Fishman's Pulmonary Diseases and Disorders (4th ed.)*. McGraw-Hill. ISBN 0-07-145739-9
- KIM, S., S. K. BALAKRISHNAN and D. S. GROSS, 2011 p53 Interacts with RNA polymerase II through its core domain and impairs Pol II processivity in vivo. *PloS one* 6: e22183.
- LARSEN, S., T. YOKOCHI, E. ISOGAI, Y. NAKAMURA, T. OZAKI *et al.*, 2010 LMO3 interacts with p53 and inhibits its transcriptional activity. *Biochemical and biophysical research communications* 392: 252-257.
- LAZAR, V. E. A., 1993 Simple sequence repeat polymorphism within the p53 gene. *Oncogene* 8: 1703-1705.
- LEROY, B., J. L. FOURNIER, C. ISHIOKA, P. MONTI, A. INGA *et al.*, 2013 The TP53 website: an integrative resource centre for the TP53 mutation database and TP53 mutant analysis. *Nucleic acids research* 41: D962-969.
- LEVINE, A. J., and M. OREN, 2009 The first 30 years of p53: growing ever more complex. *Nature reviews. Cancer* 9: 749-758.
- LINARES, L., A. HENGSTERMANN, A. CIECHANOVER and S. M. MULLER S, 2003 HdmX stimulates Hdm2-mediated ubiquitination and degradation of p53. *ProcNatAcadSci USA* **100**: 12009–12014.
- LIU, X., L. F. SEMPERE, Y. GUO, M. KORC, S. KAUPPINEN *et al.*, 2011 Involvement of microRNAs in lung cancer biology and therapy. *Translational research : the journal of laboratory and clinical medicine* 157: 200-208.
- LIU, Y., S. E. ELF, Y. MIYATA, G. SASHIDA, G. HUANG *et al.*, 2009 p53 regulates hematopoietic stem cell quiescence. *Cell stem cell* 4: 37-48.
- LOHMANN, D., B. PUTZ, U. REICH, J. BOHM, H. PRAUER *et al.*, 1993 Mutational spectrum of the p53 gene in human small-cell lung cancer and relationship to clinicopathological data. *The American journal of pathology* 142: 907-915.

- MANFREDI, J. J., 2010 The Mdm2-p53 relationship evolves: Mdm2 swings both ways as an oncogene and a tumor suppressor. *Genes & development* 24: 1580-1589.
- MANFREDI, L. E. G. A. J. J., 2006 The p53 tumor suppressor participates in multiple cell cycle checkpoints. *Journal of Cellular Physiology* 209: 13–20.
- MARKOWITZ, S. D., L. MYEROFF, M. J. COOPER, J. TRAICOFF, M. KOCHERA *et al.*, 1994 A benign cultured colon adenoma bears three genetically altered colon cancer oncogenes, but progresses to tumorigenicity and transforming growth factor-beta independence without inactivating the p53 tumor suppressor gene. *The Journal of clinical investigation* 93: 1005-1013.
- MATHIEU, M. E., C. SAUCOURT, V. MOURNETAS, X. GAUTHEREAU, N. THEZE *et al.*, 2012 LIF-dependent signaling: new pieces in the Lego. *Stem cell reviews* 8: 1-15.
- MARIN, M., J. CA. and B. LA, 2000 A common polymorphism acts as an intragenic modifier of mutant P53 behaviour. *Nat Genet* 25.
- MATLASHEWSKI, G. J., S. TUCK, D. PIM, P. LAMB, J. SCHNEIDER *et al.*, 1987 Primary structure polymorphism at amino acid residue 72 of human p53. *Molecular and cellular biology* 7: 961-963.
- MC EVOY, J., A. ULYANOV, R. BRENNAN, G. WU, S. POUNDS *et al.*, 2012 Analysis of MDM2 and MDM4 single nucleotide polymorphisms, mRNA splicing and protein expression in retinoblastoma. *PloS one* 7: e42739.
- MILLER DP, L. G., VIVO ID, ET AL, 2002 Combinations of the variant genotypes of GSTP1, GSTM1, and p53 are associated with an increased lung cancer risk. *Cancer Res* 62: 2819–2823.
- MIRZAYANS, R., B. ANDRAIS, A. SCOTT and D. MURRAY, 2012 New insights into p53 signaling and cancer cell response to DNA damage: implications for cancer therapy. *Journal of biomedicine & biotechnology* 2012: 170325.
- MIYASHITA, T., S. K., M. KRAJEWSKA, 1994 Tumor suppressor p53 is a regulator of bcl-2 and bax gene expression in vitro and in vivo. *Oncogene* 9: 1799–1805.
- MURATA, M., TAGAWA, M., KIMURA, M., KIMURA, H., WATANABE, S., AND SAISHO, H. , 1996 Analysis of a germ line polymorphism of the p53 gene in lung cancer patients; discrete results with smoking history. *Carcinogenesis (Lond.)* 17: 261–264.
- NAN, H., A. A. QURESHI, D. J. HUNTER and J. HAN, 2008 Interaction between p53 codon 72 polymorphism and melanocortin 1 receptor variants on suntan response and cutaneous melanoma risk. *The British journal of dermatology* 159: 314-321.
- NEERAJ JAIN, P. V. S., MD; SURESH HEDAU, PhD; SURESH KUMAR, MD; MRADUL K. DAGA, MD; RICHADWAN, MD; NANDAGUDI S. MURTHY, PhD; SYED A. HUSAIN, PhD; AND BHUDEV C. DAS, PhD, 2005 Infection of Human Papillomavirus Type 18 and p53 Codon 72 Polymorphism in Lung Cancer Patients From India. *CHEST* 128: 3999–4007.
- NEWHAM, G. M., M. CONRON, S. MCLACHLAN, A. DOBROVIC, H. DO *et al.*, 2011 Integrated mutation, copy number and expression profiling in resectable non-small cell lung cancer. *BMC cancer* 11: 93.
- OLIVIER, M., M. HOLLSTEIN and P. HAINAUT, 2010 TP53 mutations in human cancers: origins, consequences, and clinical use. *Cold Spring Harbor perspectives in biology* 2: a001008.
- OLTVAI, Z. N., C. L. M., AND S. J. KORSMEYER, 1993 Bcl-2 heterodimerizes in vivo with a conserved homolog, bax, that accelerates programmed cell death. *Cell cycle* 74: 609–619.
- OZBEY, U., H. YUCE, M. NAMLI and T. ELKIRAN, 2011 Investigation of Differences in P53 Gene Polymorphisms between Schizophrenia and Lung Cancer Patients in the Turkish Population. *Genetics research international* 2011: 483851.
- PANDIMA, D. K., B. SIVAMARUTHI, P. V. KIRUTHIGA and S. K. PANDIAN, 2010 Study of p53 codon 72 polymorphism and codon 249 mutations in Southern India in relation to age, alcohol drinking and smoking habits. *Human and Experimental Toxicology* 29: 451–458.
- PAPADAKIS, E. D., N. SOULITZIS and D. A. SPANDIDOS, 2002 Association of p53 codon 72 polymorphism with advanced lung cancer: the Arg allele is preferentially retained in tumours arising in Arg/Pro germline heterozygotes. *British journal of cancer* 87: 1013-1018.
- PARK, K. S., M. C. LIANG, D. M. RAISER, R. ZAMPONI, R. R. ROACH *et al.*, 2011 Characterization of the cell of origin for small cell lung cancer. *Cell cycle* 10: 2806-2815.
- PARKIN DM, L. E., MUIR CS, 2001 Global cancer statistics in the year 2000. *Lancet Oncol* 2.

- PARKIN DM, L. E., MUIR CS, 1988 Estimates of the worldwide frequency of sixteen major cancers in 1980. *Int J Cancer* 41: 184- 197.
- PARKIN DM, P. P., FERLAY J., 1993 Jun Estimates of the worldwide incidence of eighteen major cancers in 1985. *Int J Cancer* 54: 594-606.
- PARKIN DM, P. P., FERLAY J., 1999 Mar Estimates of the worldwide incidence of 25 major cancers in 1990 *Int J Cancer* 80: 827-841.
- PARKIN, D. M., J. STJERNWARD and C. S. MUIR, 1984 Estimates of the worldwide frequency of twelve major cancers. *Bulletin of the World Health Organization* 62: 163-182.
- PATEL JD, B. P., KRIS M, 2004 Lung cancer in US women: a contemporary epidemic. *JAMA* 291: 1763–1768.
- PFEIFER, G. P., and A. BESARATINIA, 2009 Mutational spectra of human cancer. *Human genetics* 125: 493-506.
- PLATI, J., O. BUCUR and R. KHOSRAVI-FAR, 2011 Apoptotic cell signaling in cancer progression and therapy. *Integrative biology : quantitative biosciences from nano to macro* 3: 279-296.
- PUNTE, X. S., G. VELASCO, A. GUTIERREZ-FERNANDEZ, J. BERTRANPETIT, M. C. KING *et al.*, 2006 Comparative analysis of cancer genes in the human and chimpanzee genomes. *BMC genomics* 7: 15.
- RAHMAN, S. M., A. L. GONZALEZ, M. LI, E. H. SEELEY, L. J. ZIMMERMAN *et al.*, 2011 Lung cancer diagnosis from proteomic analysis of preinvasive lesions. *Cancer research* 71: 3009-3017.
- REED, T. M. A. J. C., 1995 Tumor suppressor p53 is a direct transcriptional activator of the human bax gene. *Cell cycle* 80: 293–299.
- RICKS-SANTI, L., T. MASON, V. APPREY, C. AHAGHOTU, A. MCLAUCHLIN *et al.*, 2010 p53 Pro72Arg polymorphism and prostate cancer in men of African descent. *The Prostate* 70: 1739-1745.
- RIVLIN, N., R. BROSH, M. OREN and V. ROTTER, 2011 Mutations in the p53 Tumor Suppressor Gene: Important Milestones at the Various Steps of Tumorigenesis. *Genes & cancer* 2: 466-474.
- ROBLES, A. I., and C. C. HARRIS, 2010 Clinical outcomes and correlates of TP53 mutations and cancer. *Cold Spring Harbor perspectives in biology* 2: a001016.
- ROTH, J., KOCH, P., CONTENTE, A. & DOBBELSTEIN, M. TUMOR-DERIVED MUTATIONS WITHIN THE DNA-BINDING DOMAIN OF P53 THAT PHENOTYPICALLY RESEMBLE THE DELETION OF THE PROLINE-RICH DOMAIN. , 2000 Tumor-derived mutations within the DNA-binding domain of p53 that phenotypically resemble the deletion of the proline-rich domain. *Oncogene* 19: 1834–1842
- ROTTER, R. B., 2009 When mutants gain new powers. *Nature Reviews Cancer* 9: 701–713.
- SAKAMURO, D., S. P., E. WHITE and G. PRENDERGAST, 1997 The polyproline region of P53 is required to activate apoptosis but not growth arrest. *Oncogene* 15: :887-898
- SAMET, J. M., E. AVILA-TANG, P. BOFFETTA, L. M. HANNAN, S. OLIVO-MARSTON *et al.*, 2009 Lung cancer in never smokers: clinical epidemiology and environmental risk factors. *Clinical cancer research : an official journal of the American Association for Cancer Research* 15: 5626-5645.
- SCHMID, K., T. KUWERT and H. DREXLER, 2010 Radon in indoor spaces: an underestimated risk factor for lung cancer in environmental medicine. *Deutsches Arzteblatt international* 107: 181-186.
- SCHNEIDER-STOCK, R., C. BOLTZE, B. PETERS, R. SZIBOR, O. LANDT *et al.*, 2004 Selective loss of codon 72 proline p53 and frequent mutational inactivation of the retained arginine allele in colorectal cancer. *Neoplasia* 6: 529-535.
- SHI, H., S. J. TAN, H. ZHONG, W. HU, A. LEVINE *et al.*, 2009 Winter temperature and UV are tightly linked to genetic changes in the p53 tumor suppressor pathway in Eastern Asia. *American journal of human genetics* 84: 534-541.
- SOPORI, M., 2002 Effects of cigarette smoke on the immune system. *Nature reviews. Immunology* 2: 372-377.
- STEGH, A. H., 2012 Targeting the p53 signaling pathway in cancer therapy - the promises, challenges and perils. *Expert opinion on therapeutic targets* 16: 67-83.
- STEWART, D. J., 2010 Tumor and host factors that may limit efficacy of chemotherapy in non-small cell and small cell lung cancer. *Critical reviews in oncology/hematology* 75: 173-234.
- SZYMANOWSKA., A., EWAJASSEM., RAFALDZIADZIUSZKO., J. L. AKE BORG, GRAZYNAKOBIERSKA-GULIDA. *et al.*, 2006 Increased risk of non-small cell lung cancer and frequency of somatic TP53 gene mutations in Pro72 carriers of TP53 Arg72Pro polymorphism. *Lung Cancer* 52: 9–14.

- TAGAWA M, M. M., KIMURA H, 1998 Prognostic value of mutations and a germ line polymorphism of the p53 gene in non-small cell lung carcinoma: association with clinicopathological features. *Cancer Lett* 128: 93–99.
- TAYLOR, R., F. NAJAFI and A. DOBSON, 2007 Meta-analysis of studies of passive smoking and lung cancer: effects of study type and continent. *International journal of epidemiology* 36: 1048-1059.
- TOLEDO, F., C. J. LEE, K. A. KRUMMEL, L. W. RODEWALD, C. W. LIU *et al.*, 2007 Mouse mutants reveal that putative protein interaction sites in the p53 proline-rich domain are dispensable for tumor suppression. *Molecular and cellular biology* 27: 1425-1432.
- TOLEDO, F., and G. WAHL, 2006 Regulating the p53 pathway: in vitro hypotheses, in vivo veritas. *Nat Rev Cancer* 6: 909–923.
- VENOT, C., M. MARATRAT, C. DUREUIL, E. CONSEILLER, L. BRACCO *et al.*, 1998 The requirement for the p53 proline-rich functional domain for mediation of apoptosis is correlated with specific PIG3 gene transactivation and with transcriptional repression. *The EMBO journal* 17: 4668-4679.
- VOGELSTEIN, B., D. L., AND A. J. LEVINE, 2000 Surfing the p53 network. *Nature cell biology* 16: 307–310.
- W. XUE, L. Z., C. MIETHING, 2007 Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature reviews. Cancer* 445: 656–660.
- WALERYCH, D., M. NAPOLI, L. COLLAVIN and G. DEL SAL, 2012 The rebel angel: mutant p53 as the driving oncogene in breast cancer. *Carcinogenesis* 33: 2007-2017.
- WANG, Y., Y. A. SUH, M. Y. FULLER, J. G. JACKSON, S. XIONG *et al.*, 2011 Restoring expression of wild-type p53 suppresses tumor growth but does not cause tumor regression in mice with a p53 missense mutation. *The Journal of clinical investigation* 121: 893-904.
- WANG YC, L. H., CHEN SK, ET AL, 1999 Prognostic significance of p53 codon 72 polymorphism in lung carcinomas. *Eur J Cancer* 35: 226–230.
- WANG YC, L. H., CHEN SK, ET AL., 1999 Prognostic significance of p53 codon 72 polymorphism in lung carcinomas *Eur J Cancer* 35: 226–230.
- WESTON, A., and J. H. GODBOLD, 1997 Polymorphisms of H-ras-1 and p53 in breast cancer and lung cancer: a meta-analysis. *Environmental health perspectives* 105 Suppl 4: 919-926.
- WESTON, A., LING-CAWLEY, H. M., CAPORASO, N. E., BOWMAN, E. D., HOOVER, R. N., TRUMP, B. F., AND HARRIS, C. C. , 1994 Determination of the allelic frequencies of an L-myc and a p53 polymorphism in human lung cancer. *Carcinogenesis (Lond.)* 15: 583–587.
- WESTON A, P. L., FORRESTER K, HOOVER RN, TRUMP BF, HARRIS CC 481-483, 1992a Allelic frequency of a p53 polymorphism in human lung cancer. *Cancer Epidemiol Biomarkers Prev* 1: 481-483.
- WESTON A, P. L., FORRESTER K, HOOVER RN, TRUMP BF, HARRIS CC, ET AL. . , 1992b Allelic frequency of a p53 polymorphism in human lung cancer. *Cancer Epidemiol Biomarkers Prev* 1.
- WHO, 2008 *World Cancer Report*. World Health Organization.
- WHO, 2004 *World Cancer Report*. World Health Organization (country office of bangladesh).
- XIFENG WU, H., CHRISTOPHER L, AMOS, SANJAY SHETE, NIMISHAMAKAN, WAUN K, HONG, FRED F, KADLUBAR, MARGARET R, SPITZ. . 2002 MAY 1:94(9):, 2002 P53 Genotypes and Haplotypes associated With Lung Cancer Susceptibility and Ethnicity. *Journal of the National Cancer Institute* 94: 681- 690.
- YI-CHING WANG, C.-Y. C., SHIN-KUANG CHEN, YUAN-YEN CHANG, AND PINPIN LIN, 1999 p53 Codon 72 Polymorphism in Taiwanese Lung Cancer Patients: Association with Lung Cancer Susceptibility and Prognosis. *Clin Cancer Res* 5: 129-134.
- YING CHUAN HU, M. P. M., AND STEVEN A. A. HRENDT, 2005 The p53 codon 72 Proline is associated with p53 gene mutations in Non- small cell lung cancer. *Clin Cancer Res* 11.